organic papers

Acta Crystallographica Section E Structure Reports Online

ISSN 1600-5368

Kiriko Kurokawa,^a Nobuko Kanehisa,^b* Masaaki Sawa,^a Hirosato Kondo^a and Yasushi Kai^b

^aChemistry Group, R&D Laboratories, Research and Development Division, Nippon Organon KK, 1-5-90 Tomobuchi-cho, Miyakojima-ku, Osaka 534-0016, Japan, and ^bDepartment of Materials Chemistry, Graduate School of Engineering, Osaka University, Suita, Osaka 565-0871, Japan

Correspondence e-mail: kanehisa@ap.chem.eng.osaka-u.ac.jp

Key indicators

Single-crystal X-ray study T = 294 KMean $\sigma(\text{C}-\text{C}) = 0.004 \text{ Å}$ R factor = 0.041 wR factor = 0.102 Data-to-parameter ratio = 10.6

For details of how these key indicators were automatically derived from the article, see http://journals.iucr.org/e.

(+)-(3*R*)-*N*-Hydroxy-2-[(*S*)-ethoxy(4-methoxyphenyl)phosphoryl]-1,2,3,4-tetrahydroisoquinoline-3-carboxamide

The title compound, $C_{19}H_{23}N_2O_5P$, is one of two diastereomers of an arylphosphonamide hydroxamate. The absolute configuration has been determined to establish the relationship between the stereochemistry at the P atom and the activity in inhibition of matrix metalloproteinases. Received 20 August 2001 Accepted 28 August 2001 Online 11 September 2001

Comment

Matrix metalloproteinases (MMPs) are a family of zincmetalloproteinases involved in the extracellular matrix degradation (Woessner, 1991). So far, more than 20 MMPs have been isolated and they are implicated in the pathological processes of a variety of diseases associated with excessive degradation of the extracellular matrix, such as tumor metastasis (Yip *et al.*, 1999), rheumatoid arthritis and osteoarthritis (Clark *et al.*, 2000), multiple sclerosis (Gijbels *et al.*, 1994). Therefore, a number of laboratories have vigorously conducted the development of MMP inhibitors as curatives of these diseases (Whittaker *et al.*, 1999).



inactive isomer (I)

Recently, our group has reported that a phosphonamidebased hydroxamate showed potent inhibitory activities against MMP-1, -3 and -9 (Sawa *et al.*, 2001). It was found that only one of the two diastereomers resulting from the chirality at the P atom showed potent inhibition against MMPs [active isomer: (II)], while the other isomer was inactive [inactive isomer: (I)]. This result suggested that the stereochemistry at the P atom was very important for the inhibition of the enzymes.





© 2001 International Union of Crystallography Printed in Great Britain – all rights reserved Therefore, the establishment of the absolute configuration of the P atom would be a great help in studying the interactions of the

inhibitor in the active sites of MMPs. Moreover, it seems to be worthwhile information for the design of protease inhibitors. The inactive isomer (I) was used for the X-ray analysis instead of the active isomer (II), because (I) could be much more easily crystallized than (II).

As shown in Fig. 1, the configuration of the P atom of the inactive isomer (I) was determined to be S. The configuration of one of the two chiral centers, the α -carbon of the hydroxamate group, was already known, because both (I) and (II) were synthesized from the known *R*-form of the starting material. The resultant configuration of the α -carbon, C2, is consistent with this stereochemistry (*R* configuration). Hence, the P atom of the active isomer (II) is in the *R* configuration. Recently, Pikul *et al.* (1999) reported that a phosphinamide-based hydroxamate (III) having an *R* configuration at the P atom exhibited inhibitory activities against MMPs, while the S isomer at the P atom was a poor inhibitor. As expected, the stereochemistry of the P atom of (II) was consistent with that of (III).



The structural data of (I) reveal that the non-aromatic portion of the tetrahydroisoquinoline unit adopts a half-chair conformation. The torsion angles C2-C3-C9-C8 and N1-C1-C8-C9 are 29.7 (4) and -9.5 (4)°, respectively. Comparison of the phosphonamide portion of (I) with the corresponding phosphinamide portion of (III) reveals structural differences as follows. The bond lengths around the P atom of (I), P1-O3, P1-N1 and P1-C13 are similar to the



Figure 1

The molecular structure of (I) showing the atom-numbering scheme. Displacement ellipsoids are drawn at the 40% probability level.

corresponding bond lengths of (III), within a range of 0.02 Å. On the other hand, the bond angles around the P atom of (I) are different from those of (III). The O3-P1-C13 bond angle of (I) is larger than the corresponding bond angle of (III) [114.0 (1) and 108.3°, respectively], whereas the O4-P1-C13 and N1-P1-C13 bond angles are smaller than those of (III) [O4-P1-C13 105.5 (1)° for (I) and 108.3° for (III); N1-P1-C13 107.9 (1)° for (I) and 110.5° for (III)]. The P1-C13 bond of (I) leans further towards the N1-P1-O4 plane compared to the situation in (III).

Experimental

The synthesis of the title compound is reported elsewhere (Sawa *et al.*, 2001). Crystals of (I) were obtained by recrystallization from EtOH at 293 K. The melting point of (I) is 440.0–440.5 K. The optical rotation of (I), $[\alpha]_D^{-20}$, is +68.43° (*c* 1.02, MeOH) and that of (II) is +18.48° (*c* 1.10, MeOH).

Crystal data

 $C_{19}H_{23}N_2O_5P$ Cu Ka radiation $M_r = 390.37$ Cell parameters from 21 Orthorhombic, P212121 reflections a = 10.156 (2) Å $\theta = 27.5 - 30.0^{\circ}$ $\mu = 1.57~\mathrm{mm}^{-1}$ b = 19.733(1) Å c = 9.486(1) Å T = 294.2 KV = 1901.1 (3) Å² Prismatic, colorless $0.20 \times 0.10 \times 0.10 \ \mathrm{mm}$ Z = 4 $D_x = 1.364 \text{ Mg m}^{-3}$

 $\begin{aligned} R_{\rm int} &= 0.016\\ \theta_{\rm max} &= 62.0^\circ \end{aligned}$

 $h = -9 \rightarrow 11$

 $k = -22 \rightarrow 22$

 $l = -10 \rightarrow 10$

3 standard reflections

every 150 reflections

intensity decay: 3.7%

Data collection

Rigaku AFC-5*R* diffractometer ω -2 θ scans Absorption correction: ψ scan (North *et al.*, 1968)

 $T_{\min} = 0.757, T_{\max} = 0.854$ 3303 measured reflections 3303 independent reflections 2991 reflections with $F^2 > 2\sigma(F^2)$

Refinement

 $w = 1/[\sigma^{2}(F_{o}^{2}) + \{0.019[Max(F_{o}^{2},0) + 2F_{c}^{2}]/3\}^{2}]$ Refinement on F^2 $R[F^2 > 2\sigma(F^2)] = 0.041$ $wR(F^2) = 0.102$ $(\Delta/\sigma)_{\rm max} = 0.013$ $\Delta \rho_{\rm max} = 0.17 \text{ e } \text{\AA}^{-3}$ S=1.65 $\Delta \rho_{\rm min} = -0.20 \ {\rm e} \ {\rm \AA}^{-3}$ 3303 reflections Absolute structure: Flack (1983), 313 parameters H atoms treated by a mixture of 1571 Friedel pairs Flack parameter = 0.01 (3) independent and constrained refinement

Table 1

Selected geometric parameters (Å, °).

P1-O3	1.477 (2)	P1-N1	1.636 (3)
P1-O4	1.584 (2)	P1-C13	1.793 (3)
O3-P1-O4	112.6 (1)	O4-P1-N1	104.5 (1)
O3-P1-N1	111.7 (1)	O4-P1-C13	105.5 (1)
O3-P1-C13	114.0 (1)	N1-P1-C13	107.9 (1)
$N_{1} = C_{1} = C_{2} = C_{2}$	0.5 (4)	C^{2} C^{2} C^{0} C^{0}	20.7 (4)
NI-CI-Co-C9	-9.5 (4)	02=03=09=08	29.7 (4)

Table 2Hydrogen-bonding geometry (Å, °).

$D - H \cdot \cdot \cdot A$	D-H	$H \cdots A$	$D \cdots A$	$D - H \cdots A$
$\begin{array}{c} O2 - H11 \cdots O1 \\ N2 - H10 \cdots N1 \\ N2 - H10 \cdots O1^{i} \end{array}$	0.90 (4)	2.44 (3)	2.701 (2)	97 (2)
	0.88 (3)	2.46 (2)	2.703 (2)	96 (1)
	0.88 (3)	2.12 (2)	2.951 (2)	158 (2)

Symmetry code: (i) $\frac{1}{2} + x, \frac{3}{2} - y, 1 - z$.

The maximum θ was restricted to 62° by the diffractometer geometry. The methyl H atoms on C12 and C19 were placed at geometrically idealized positions (C-H = 0.95 Å) and not refined. All other H atoms were found from the difference Fourier maps and refined isotropically. The C-H, N-H and O-H bond lengths are 0.90 (3)–1.07 (4), 0.88 (3) and 0.90 (4) Å, respectively.

Data collection: *Rigaku/AFC Diffractometer Control Software* (Rigaku, 1998); cell refinement: *Rigaku/AFC Diffractometer Control Software*; data reduction: *teXsan* (Molecular Structure Corporation, 1999); program(s) used to solve structure: *SIR*92 (Altomare *et al.*, 1994); program(s) used to refine structure: *teXsan*; molecular graphics: *ORTEP*II (Johnson, 1976); software used to prepare material for publication: *teXsan*.

References

- Altomare, A., Cascarano, G., Giacovazzo, C., Guagliardi, A., Burla, M. C., Polidori, G. & Camalli, M. (1994). J. Appl. Cryst. 27, 435.
- Clark, I. M., Rowan, A. D. & Cawston, T. E. (2000). Curr. Opin. Anti-Inflammat. Immunomodulat. Invest. Drugs, 2, 16–25.
- Flack, H. D. (1983). Acta Cryst. A39, 876-881.
- Gijbels, K., Galardy, R. E. & Steinman, L. (1994). J. Clin. Invest. 94, 2177-2182.
- Johnson, C. K. (1976). ORTEPII. Report ORNL-5138. Oak Ridge National Laboratory, Tennessee, USA.
- Molecular Structure Corporation (1999). *teXsan*. Version 1.10. MSC, 9009 New Trails Drive, The Woodlands, TX 77381–5209, USA.
- North, A. C. T., Phillips, D. C. & Mathews, F. S. (1968). Acta Cryst. A24, 351–359.

Pikul, S., Dunham, K. L. M., Almstead, N. G., De, B., Natchus, M. G., Anastasio, M. V., McPhail, S. J., Snider, C. E., Taiwo, Y. O., Chen, L., Dunaway, C. M., Gu, F. & Mieling, G. E. (1999). J. Med. Chem. 42, 87–94.

Rigaku (1998). Rigaku/AFC Diffractometer Control Software. Rigaku Corporation, Tokyo, Japan.

Sawa, M., Kiyoi, T., Kurokawa, K., Kumihara, H., Yamamoto, M., Miyasaka, T., Ito, Y., Hirayama, R., Inoue, T., Kirii, Y., Nishiwaki, E., Ohmoto, H., Maeda, Y., Ishibushi, E., Inoue, Y., Yoshino, K. & Kondo, H. (2001). J. Med. Chem.. Submitted.

Whittaker, M., Floyd, C. D., Brown, P. & Gearing, A. J. H. (1999). Chem. Rev. 99, 2735–2776.

Woessner, J. F. (1991). FASEB J. 5, 2145-2154.

Yip, D., Ahmad, A., Karapetis, C. S., Hawkins, C. A. & Harper, P. G. (1999). *Invest. New Drugs*, **17**, 387–399.